Expression of Matrix Metalloproteinase 1 (MMP1) in HepatoCellular Carcinoma (HCC): Immunohistochemical and Biochemical Studies
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Abstract
Background: The present study aimed to evaluate the expression of Matrix Metalloproteinase-1 (MMP1) in HepatoCellular Carcinoma (HCC) by using the Immunohistochemical technique, which allows us to integrate the biological aspects of this enzymatic expression in the morphological context of HCCs.

Material and Methods: The study was performed on 70 subjects from out and in patients of Tropical medicine Department, Thiodor Billhars Institute during the period from January 2011 until June 2012. The present study included 60 patients with chronic hepatitis C who had undergone liver biopsy. They consisted of 42 men and 28 women with ages ranging from 36 to 66 years. The diagnosis of chronic hepatitis C was made on the basis of positivity for anti-HCV (by the second generation ELISA), and confirmed by HCV-RNA reverse transcription polymerase chain reaction (RT-PCR). Patients were divided into four groups:
- Group I: included 10 normal persons with no history of liver disease with normal liver enzymes and free ultrasonographic finding as normal control. It included 6 males, 4 females, with ages ranging from 34 to 48 years.
- Group II: included 20 HCV infected patients without cirrhotic changes. It included 11 males, 9 females, with ages ranging from 39 to 53 years.
- Group III: included 20 HCV infected patients with liver cirrhosis, 12 males, 8 females, with ages ranging from 48-63 years.
- Group IV: included 20 HCV infected patients with HCC, 16 males, 4 females, with ages ranging from 53-64 years.

Results: Blood Picture, (Hb, WBCs, RBCs, Plts, PC and ESR). Liver Function Test (ALT, AST, ALB, GGT, ALP, T. BIL and D. BIL). Matrix Metalloprotenase 1(MMP1) Measurements: Serum MMP1. Histopathological investigation Including histopathological changes in the liver tissue.

Conclusion: our results suggest that MMP-1 is overexpressed in a large proportion of patients with HCC and the high expression level of protein correlated with the disease progression and poor clinical outcome in HCC. Furthermore, MMP-1 high expression proved to be a risk factor for tumor recurrence and independent molecular marker of prognosis in HCC and may become a novel target in the strategies for the prediction of tumor progression and prognosis of this disease.

Key words: Patients, HCV, Cirrhosis, HCC, and MMP-1

Introduction: Hepatocellular carcinoma (HCC) is the fifth most common cancer and sixth leading cause of death among cancers worldwide (1), and HCC is a heterogeneous group in terms of biological behavior and molecular profiles (2). Recent studies support that tumors may be initiated and maintained by a small population of cells that have stem-like features, and this highly tumorigenic cell subset within the tumor bulk has been considered as cancer stem cell (3). Invasion is a characteristic feature of HCC, it frequently shows early invasion into blood vessels as well as intrahepatic metastasis and later shows extrahepatic metastasis.

Tommaso. (4), reported that, Hepatocellular carcinoma (HCC) is a common form of cancer that arises from hepatocytes and whose risk may be affected by several known environmental factors, including hepatitis viruses, alcohol, cigarette smoking, and others. Rare monogenic syndromes, such as alpha1-antitrypsin deficiency, glycogen storage disease type I, hemochromatosis, acute intermittent and cutaneous porphyria, as well as hereditary tyrosinemia type I are associated with a high risk of HCC. Several common conditions or diseases inherited as polygenic traits e.g. autoimmune hepatitis, type 2 diabetes, a family history of HCC, hypothyroidism, and non-alcoholic steatohepatitis also show an increased risk of HCC compared to the general population. Hepatitis C infection
remains the greatest risk factor for the development of HCC. Approximately 170 million people worldwide are seropositive for anti-HCV and, of these, an estimated 127 million are chronically infected (5). Markers of HCV infection are found in 28–76% of HCC cases in Europe (with an increasing gradient from north to south) and in 80–90% of patients with HCC presenting in Japan. Methods of HCV viral transmission are complex. In the United States, HCV infection is largely associated with intravenous drug use and sexual contact, which account for approximately 85% of infections (6). In 1978, cirrhosis was defined by the World Health Organization as a diffuse process characterized by fibrosis and the conversion of normal liver architecture into structurally abnormal nodules (7). The most common cause of hepatic fibrosis is alcohol abuse, which leads to repeated liver injury/damage and degeneration of liver parenchymal tissue. Apart from alcohol abuse, other factors have the potential to induce hepatic fibrogenesis. Cirrhosis and chronic repetitive hepatic injury, as are manifest in chronic active hepatitis, are associated with the majority (80%) of HCC cases and represent a major underlying factor predisposing the development of HCC (8).

In 2004, Matsumaga et al. (9) demonstrated that the expression of MMP-1 in most of the HCC tissues was equal or low compared with those in the surrounding non-tumor tissues, although mixed expression pattern was recognized in some HCC tissues. The difference of MMP-1 expression was not related with the histological differentiation of HCC and the condition of non-cancerous area. These findings suggested little association of the clinicopathological findings of HCC with the histological expression of MMP-1. However, in 2009, Altadill et al. (10) reported that MMP-1 is mainly expressed by stromal cells of HCC tissues. A positive correlation between MMP-1 expression and larger size tumors was found. Moreover, they also found that all HCC patients showing elevated MMP-1 expression in stromal cells presented a poor prognosis.

Material and Methods:
I-Materials:
The study was performed on 70 subjects from out and in patients of Tropical medicine Department, Thiodor Billhars Institute during the period from January 2011 until June 2012. The present study included 60 patients with chronic hepatitis C who had undergone liver biopsy. They consisted of 42 men and 28 women with ages ranging from 36 to 66 years. The diagnosis of chronic hepatitis C was made on the basis of positivity for anti-HCV (by the second generation ELISA), and confirmed by HCV-RNA reverse transcription polymerase chain reaction (RT-PCR).

Patients with HBV infection or auto antibodies (antinuclear antibody, anti-smooth muscle antibody, and antimitochondrial antibody, or history of alcohol abuse were excluded from the study.

Patients were divided into four groups:
Group I: included 10 normal persons with no history of liver disease with normal liver enzymes and free ultrasonographic finding as normal control. It included 6 males, 4 females, with ages ranging from 34 to 48 years.
Group II: included 20 HCV infected patients without cirrhotic changes. It included 11 males, 9 females, with ages ranging from 39 to 53 years.
Group III: included 20 HCV infected patients with liver cirrhosis, 12 males, 8 females, with ages ranging from 48-63 years.
Group IV: included 20 HCV infected patients with HCC, 16 males, 4 females, with ages ranging from 53-64 years.

A detailed history and physical examination of the patients were carried out with special emphasis on history of schistosomiasis, prior parenteral therapy, infective hepatitis and jaundice or other signs of liver cell failure. Complete clinical examination, which includes the manifestations of hepatitis and liver cell failure such as jaundice, hepatomegaly, tenderness in the right hypochondrium, ascites, splenomegaly, lower limb edema as well as abdominal ultrasonography was also done side by side with routine laboratory investigations.

Biochemical and Serological Tests: Ten milliliters of fasted venous blood (6 Hours of fasting) were taken from each subject participating in the study.1.0 ml of blood added into EDTA tubes for determination of Hemoglobin, RBCs, WBCs, Platelets and Erythrocytes Sedimentation Rate (ESR) and the rest of the blood was left to clot. Serum were separated by centrifugation and stored at -20°C for analysis of:
Markers of Hepatitis Virus: Hepatitis B surface antigen (HBsAg), hepatitis B core antibody
(HBCAb), hepatitis C virus antibody (HCV-Ab), and hepatitis C virus RNA (HCV-RNA). Markers of Hepatitis Virus: HCV antibodies were detected using a third generation enzyme-linked immunosorbent assay (Sorin Biomedica Diagnostics, Italy), (11). Serological assay for HBV markers (HbsAg and anti-HBc) were performed by a direct noncompetitive sandwich assay (DiaSorin, Italy) based on ELISA technique (12).

**Blood Picture:** Hemoglobin (Hb), white blood cell counts (WBCs), red blood cell counts (RBCs), and Erythrocytes Sedimentation Rate (ESR).

Blood Picture was done on Coulter Counter T890, (Coulter Counter, Harpenden, UK) (13).

**Liver Function Test:** Alanine aminotransferase (ALT), aspartate aminotransferase (ASAT), albumin (ALB), γ-glutamyl Transferase (GGT), Alkaline Phosphatase (ALP), Total bilirubin (T. BIL) and Direct bilirubin (D. BIL). Liver function tests were performed using a Beckman Auto-analyzer (Synchron CX4, USA). A diazotization method used for determination of serum total bilirubin (14). Activities of ALT and AST were measured by the enzyme rate method (15). Albumin was determined according to Pinnell and Northam, (16). Prothrombin time was determined using standard thromboplastin method (17).

**Matrix Metalloproteinase 1(MMP1) Measurements:** Serum MMP1.

**Liver Biopsy:** Liver biopsy samples were obtained for diagnostic purposes percutaneously, in some cases HCC was diagnosed guided by ultrasound using a Toshiba SSA 240, an apparatus with a 3.5 MHz probe.

**II-METHODS:**

Liver biopsies will proceed in standard method to paraffin blocks and stained with:
1- H&E stain for histopathological diagnosis of cases.
2- Masson Trichrome to evaluate fibrosis.
3- Immunohistochemical stain using monoclonal antibody for Matrix Metalloproteinase 1(MMP1) according to Hsu and Raine (18).

**II.1-Immunohistochemical Procedure:**

Tissue sections from all different studied bladder lesions were immunohistochemically stained for MMP1 using the standard avidin-biotin peroxidase complex (APC) according to Hsu and Raine (18) method through the following procedures:

F. Counter stain. G. Dehydration of the sections. H. Mounting procedures.

**II.2-Histopathological invistigation:**

Including histopathological changes in the liver tissue.


Liver sections were microscopically studied to evaluate the pathological changes including portal tracts and the schistosomal granulomatous reactions.

**II.3- Biochemical parameters:**

Including liver function tests (ALAT, ASAT, ALB, GGT, ALP, T. BIL and D. BIL).

**II.4- Statistical analysis:**

Results were expressed as the mean ± SE. Data were statistically analyzed for variance and the least significant difference (LSD) using one way analysis of variance (ANOVA) according to Snedecor and Cochran, (19). SPSS version 13 was used for analysis.

**Results**

1- The results of biochemical parameters of serum liver Functions of all Patients were documented in Table (1):

**Estimation of serum liver function tests (ALAT, ASAT, ALB, GGT, ALP, T. BIL and D. BIL):**

In the present data, a high significant increase in all parameter of liver functions enzymes activity (ASAT, ALAT, ALP, γGT, T. BIL and T. BIL) and high significant decrease of albumin in HCV, Cirrhosis and MMP1 groups versus the uninfected negative control group. An amelioration in all parameters of liver functions enzymes activity of HCV as comparing to Cirrhosis as comparing to MMP1 groups.

**II. The results of Complete Blood Picture of all Patients were documented in Table (2):**

In the present data, high significant increase in all parameter of CBC (PC, WBCs, RBCs, Hb%, Platelets) and ESR in HCV, Cirrhosis and MMP1 groups versus the uninfected negative control group. An amelioration in all parameters of CBC of HCV as comparing to Cirrhosis and Cirrhosis as comparing to MMP1 groups.

**III- The results of Matrix Metalloproteinase 1(MMP1) of all Patients were documented in Table (3):**

The results demonstrated an increasing of MMP1 with a high significant differences (p<0.01) in HCC group as comparing to each of
normal control HCV and Cirrhosis groups, also increasing of MMP1 with high significant differences \((p< 0.01)\) in Cirrhosis groups as comparing to each of normal control and HCV groups. On the other hand there is increase of MMP1 in HCV group but with no significant differences as comparing to normal control group.

VII- Histopathological grading and Staging of the studied HCV, Cirrhosis and HCC groups:

The Evaluation demonstrated that, in HCV group grade I and II represent 55% and 45% respectively, while there is no grade III. On the other hand there are no grade I and II, while grade III represent 100% in Cirrhosis group. HCC grade I,II and III reaches 50%, 40% and 10% respectively as shown in (Table 4 and 5).

Histopathological Staging of the studied HCV represent 55% and 45% for the Staging I and II respectively, while there is no Staging III. On the other hand, Staging III in Cirrhosis group represent 100% and there are no Staging I and II as shown in (Table 6).

<table>
<thead>
<tr>
<th>Groups</th>
<th>ASAT (U/L)</th>
<th>sALAT (U/L)</th>
<th>ALP (U/L)</th>
<th>γGT (U/L)</th>
<th>Alb (mg/dl)</th>
<th>D. Bil</th>
<th>T. Bil</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal control</td>
<td>n=10</td>
<td>Mean ± SE</td>
<td>18.60 ± 2.01</td>
<td>21.80 ± 2.08</td>
<td>99.50 ± 8.81</td>
<td>20.00 ± 2.08</td>
<td>4.08 ± 0.39</td>
</tr>
<tr>
<td>patients with</td>
<td>n=20</td>
<td>Mean ± SE</td>
<td>a’ ± 11.38</td>
<td>a’ ± 13.25</td>
<td>a’ ± 11.18</td>
<td>a’ ± 11.05</td>
<td>a’ ± 0.36</td>
</tr>
<tr>
<td>HCV</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>patients with</td>
<td>n=20</td>
<td>Mean ± SE</td>
<td>74.80 ± 9.8</td>
<td>73.40 ± 8.3</td>
<td>178.53 ± 8.7</td>
<td>121.800 ± 19.1</td>
<td>3.12 ± .31</td>
</tr>
<tr>
<td>chronic cirrhosis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>patients with</td>
<td>n=20</td>
<td>Mean ± SE</td>
<td>90.40 ± 5.8</td>
<td>102.88 ± 10.2</td>
<td>295.68 ± 22.9</td>
<td>135.28 ± 11.9</td>
<td>2.91 ± .29</td>
</tr>
<tr>
<td>(HCC)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

- Each value represents the mean of number of patients in each group ± SE.

a= p<0.05 & a’= p< 0.01 Significant different from normal control group.
b= p<0.05 & b’= p< 0.01 Significant different from HCV Group.
c= p<0.05 & c’= p< 0.01 Significant different from Cirrhosis group.
d= p<0.05 & d’= p< 0.01 Significant different from HCC group.
Table (2): Shows the values of Complete Blood Picture in each groups and the effects of diseases on the liver.

<table>
<thead>
<tr>
<th>Groups</th>
<th>PC (Mean ± SE)</th>
<th>WBCs (Mean ± SE)</th>
<th>RBCs (Mean ± SE)</th>
<th>Hb% (Mean ± SE)</th>
<th>Platelets (Mean ± SE)</th>
<th>ESR (Mean ± SE)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal control</td>
<td>93.40 ± 1.95</td>
<td>5.60 ± .238</td>
<td>5.20 ± .075</td>
<td>14.80 ± 1.32</td>
<td>197.20 ± 16.61</td>
<td>10.800 ± 1.14</td>
</tr>
<tr>
<td>patients with HCV</td>
<td>c’ d’</td>
<td>a c d’</td>
<td>c’ d’</td>
<td>a c d’</td>
<td>a’ c d’</td>
<td>a c d’</td>
</tr>
<tr>
<td>n=15</td>
<td>82.33 ± 7.05</td>
<td>5.12 ± .405</td>
<td>4.24 ± .316</td>
<td>12.16 ± 1.18</td>
<td>144.40 ± 5.40</td>
<td>17.46 ± 1.85</td>
</tr>
<tr>
<td>patients with chronic</td>
<td>a’ b’</td>
<td>a’ b’ d’</td>
<td>a’ b’ d’</td>
<td>a’ b’ d’</td>
<td>a’ b’ d’</td>
<td>a’ b’ d’</td>
</tr>
<tr>
<td>cirrhosis n=15</td>
<td>69.466 ± 6.78</td>
<td>4.39 ± .343</td>
<td>3.94 ± .120</td>
<td>11.46 ± 0.80</td>
<td>131.40 ± 5.28</td>
<td>29.40 ± 3.01</td>
</tr>
<tr>
<td>patients with (HCC)</td>
<td>a’ b’ c’</td>
<td>a’ b’ c’</td>
<td>a’ b’ c’</td>
<td>a’ b’ c’</td>
<td>a’ b’ c’</td>
<td>a’ b’ c’</td>
</tr>
<tr>
<td>n=20</td>
<td>64.76 ± 5.59</td>
<td>3.87 ± .265</td>
<td>3.63 ± .280</td>
<td>10.23 ± 1.06</td>
<td>101.08 ± 9.32</td>
<td>54.96 ± 4.91</td>
</tr>
</tbody>
</table>

- Each value represents the mean of number of patients in each group ± SE.
  a= p<0.05 & a’= p< 0.01 Significant different from normal control group.
  b= p<0.05 & b’= p< 0.01 Significant different from HCV Group.
  c= p<0.05 & c’= p< 0.01 Significant different from Cirrhosis group.
  d= p<0.05 & d’= p< 0.01 Significant different from HCC group.

Table (3): Shows the Mean Difference of Matrix Metalloproteinase 1(MMP1).

<table>
<thead>
<tr>
<th>Groups</th>
<th>MMP1 (Mean ± SE)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1.6± .066</td>
</tr>
<tr>
<td>HCV</td>
<td>c’ d’ 2.600 ± .180</td>
</tr>
<tr>
<td>cirrhosis</td>
<td>a’ b’ 10.200 ± 1.07</td>
</tr>
<tr>
<td>(HCC)</td>
<td>a’ b’ c’ 22.360 ± 2.91</td>
</tr>
</tbody>
</table>

- Each value represents the mean of number of patients in each group ± SE.
  a’= p< 0.01 Significant different from normal control group.
  b’= p< 0.01 Significant different from HCV Group.
  c’= p< 0.01 Significant different from Cirrhosis group.
  d’= p< 0.01 Significant different from HCC group.
Table (4): Shows the Histopathological grading of the studied HCV and Cirrhosis groups:

<table>
<thead>
<tr>
<th>Groups</th>
<th>Grades</th>
<th>Number</th>
<th>Percent</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade degree</td>
<td>Number</td>
<td>Percent</td>
<td></td>
</tr>
<tr>
<td>patients with HCV n=20</td>
<td>Grade I</td>
<td>11</td>
<td>55%</td>
</tr>
<tr>
<td></td>
<td>Grade II</td>
<td>9</td>
<td>45%</td>
</tr>
<tr>
<td></td>
<td>Grade III</td>
<td>0</td>
<td>0%</td>
</tr>
<tr>
<td>patients with chronic cirrhosis n=20</td>
<td>Grade I</td>
<td>0</td>
<td>0%</td>
</tr>
<tr>
<td></td>
<td>Grade II</td>
<td>0</td>
<td>0%</td>
</tr>
<tr>
<td></td>
<td>Grade III</td>
<td>20</td>
<td>100%</td>
</tr>
</tbody>
</table>

Table (5): Shows the Histopathological grading of the studied HCC group:

<table>
<thead>
<tr>
<th>Group</th>
<th>HCC Grade</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade degree</td>
<td>Number</td>
</tr>
<tr>
<td>patients with HCC n=20</td>
<td>Grade I</td>
</tr>
<tr>
<td></td>
<td>Grade II</td>
</tr>
<tr>
<td></td>
<td>Grade III</td>
</tr>
</tbody>
</table>

Table (6): Shows the Histopathological Staging of the studied HCV and Cirrhosis groups:

<table>
<thead>
<tr>
<th>Groups</th>
<th>Stages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stages degree</td>
<td>Number</td>
</tr>
<tr>
<td>patients with HCV n=20</td>
<td>Stages I</td>
</tr>
<tr>
<td></td>
<td>Stages II</td>
</tr>
<tr>
<td></td>
<td>Stages III</td>
</tr>
<tr>
<td>patients with chronic cirrhosis n=20</td>
<td>Stages I</td>
</tr>
<tr>
<td></td>
<td>Stages II</td>
</tr>
<tr>
<td></td>
<td>Stages III</td>
</tr>
</tbody>
</table>
Figures show Histopathological grading and Staging of the studied HCV, Cirrhosis and HCC groups comparing to control group:

Figure 1: shows liver section from control case (H&E, 200).

Figure 2: shows liver section from control case showing negative expression for MMP1 protein (IHC, DAB, 200).

Figure 3: shows Chronic hepatitis C METAVIR A1F1 (H&E, 200).

Figure 4: shows Chronic hepatitis C METAVIR A1F1 showing mild expression for MMP1 in hepatocytes as cytoplasmic stain (IHC,DAB, 200).

Figure 7: shows Chronic hepatitis C METAVIR A3F4 with formation of cirrhotic nodule showing moderate positive hepatocytes expression for MMP1 as cytoplasmic stain (IHC,DAB, 200).

Figure 8: shows Hepatocellular carcinoma, grade II (H&E, 200).
Hepatocellular carcinoma (HCC) is the third most common cause of cancer-related deaths in the world (20). This tumor develops in patients with chronic liver diseases, and its etiopathogenesis includes viral infection (hepatitis B and C), alcohol, and aflatoxin B1 consumption (21). Although the diagnostic and surgical approaches have made great progress in recent years, patient survival remains unsatisfactory because of a high incidence of recurrence after hepatic resection or other types of loco-regional therapy (22). The 5-year survival rate of HCC is 25–39% following surgery (23).

The results from our study by analyzing the expression patterns of MMP-1 using immunohistochemistry assay revealed that the high MMP-1 expression levels were both associated with high fibrosis stage, inflammation grading of liver tissues, liver function activities, presence of tumor recurrence and portal vein invasion, suggesting that MMP-1 could be good diagnostic factor represent the liver states under HCV infection and followed diseases disorder. Our results in agreement with Okamoto et al. (24), who mention that, in one paper, the expression of the MMP-1 was slightly increased in HCC patients with a background of chronic hepatitis C virus (HCV)-related liver disease compared to patients with HCV-related chronic liver disease without HCC.

Our data demonstrated that, the expression patterns of MMP-1 is associated with the degree of liver diseases where the expression patterns of MMP-1 in hepatic HCC patients is a very high significant as comparing with that in hepatic cirrhosis patients that in which the expression patterns of MMP-1 is very high significant as comparing with that in hepatic HCV patients, while the expression patterns of MMP-1 in hepatic HCV elevated but without significant difference as the of collagen is slightly accumulate which degraded with MMP1 (Collaginase1). These agree with Grimm et al. (25) & Fang et al. (26), who reported that, MMP-1 expression has been described in both neoplastic and peritumoral stromal cells; however, its presence is considered more important in the zone of greatest activity corresponding to the tumor.

These results coincide with Liao et al. (27), who noticed that, The results from their study by analyzing the expression patterns of MMP-1 in 106 HCC surgical specimens using immunohistochemistry assay revealed that the high MMP-1 expression levels were both associated with high fibrosis stage, presence of tumor recurrence and portal vein invasion, suggesting that MMP-1 could be independent prognostic factors.

The present data is in parallel with Hatfield et al. (28), who represented that, MMPs,
as a family of zinc-dependent endopeptidases, are able to degrade virtually any component of the extracellular matrix. MMPs are critical for remodeling the extracellular matrix, thereby affecting cell behavior under physiologic and pathophysiologic circumstances, such as embryogenesis and cancer progression. In this family, MMP-1 initiates degradation of collagen I, which is abundant in the extracellular matrix and is essential for keratinocyte migration; several authors consider that these mechanisms facilitate tumor invasion.

The results explain that, increasing with a high significant differences of ASAT in HCV, cirrhosis and HCC groups as comparing to normal control group, while there is no significant differences in value of ASAT among the HCV, cirrhosis and HCC groups. On the other hand, the values of ALAT increasing with high significant differences in HCV, cirrhosis and HCC groups as comparing to normal control group, while there is no significant differences of values of ALAT among HCV, cirrhosis and HCC groups.

These results agree with Hann et al. (29), they observe a significant association between ALT and HCC risk in either univariate or multivariate analysis, suggesting the inability of ALT as a prospective predictor of HCC risk in HCV patients. AST exhibited a significant association with HCC risk in the univariate analysis, which disappeared after multivariate analysis adjusting all the major variables including cirrhosis. Similar observations were also noticed for ALP. These data indicated that the associations observed for AST and ALP could be potentially mediated by the presence of liver cirrhosis. (30).

Our data demonstrate that, elevation of ALP values with high significant differences in HCV, Cirrhosis and HCC groups as comparing to normal control group. In addition to there is elevation of ALP values with high significant differences in HCC in comparing to both HCV and Cirrhosis groups and there is no significant differences between HCV and Cirrhosis groups.

The data obtained are in coincidence with Giannini et al. (31) whos stated that there are two major types of serum liver enzyme level changes commonly encountered in clinical practice: hepatocellular predominance with elevated ALT and AST, and cholestatic predominance with elevated ALP and GGT. Serum ALT and AST are released from damaged hepatocytes into blood and their activities have been widely recognized as effective tools to detect liver diseases (32 & 33). Actually, ALT is the most extensively investigated serum enzyme and elevated ALT has been associated with the mortality in various liver diseases (32 & 34).

Our data also in coincidence with Aragon and Younossi (35) they noticed that, the joint analysis of GGT with other enzymes may yield additional information regarding disease risk and diagnosis. For example, elevated GGT combined with elevated ALP usually points to hepatobiliary injury, which distinguishes from ALP elevation alone resulting from bone diseases.

Our results agree with Hann et al. (29), reported that, further combined GGT with ALT, AST or ALP to determine if the combined evaluation could improve the predictive power compared to GGT alone. the combined analysis of GGT with ALP markedly increased HCC risk in patients with an elevated level of both enzymes compared to those with a normal value for both enzymes.

Our data agree with Ishizawa et al. (36), who reported that, decreased ALB indicate marked liver damage, while ascites and encephalopathy indicate liver failure and serious portal hypertension. Also agree with Zhao et al. (37) concluded that platelet count, ALP, prealbumin and GGT may be considered as supplemental factors for routine liver function scoring systems.

In the present investigation, the histopathological results indicated that, most patients were in stage 2 (28, 45.2%), while stage 1 was represented by few cases (7, 11.3%). In other words, most of the included patients (55, 88.7%) had significant fibrosis (i.e., METAVIR stages more than 1). These results indicated, therefore that, HCV infection has a rapid course of disease progression in the studied population. Similar results were reported in a similar Egyptian population by Mangoud et al. (38). These authors attributed the progressive nature of the disease to the concomitant infection with other viruses like HBV. However, in this study patients with HBV concomitant infection were
excluded. Hence, METAVIR stage 1 fibrosis was absent from the study of Mangoud et al. (39) (and METAVIR stage 2 replaced stage 1 in this study instead) due to the rapid course of disease progression caused by other concomitant infections.

These results agree with Alberti et al. (40) Staging of fibrosis is helpful in determining the duration of treatment. Lesions should be assessed histologically even in patients with persistently normal serum transaminases since advanced fibrosis has been shown in many of these patients.

In conclusion, we find that our results contribute to a better knowledge of the biological characterisation of HCC with regard to different patterns of expression of MMP1. Moreover, our findings open the possibility to design other studies on the impact of the enzymatic system of MMP1 on prognosis, therefore opening the possibility of future therapeutic targets for this frequent tumour.

References
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esophageal adenocarcinomas and is associated with positive lymph node status. J Transl Med 8:99